

Genetic nomenclature

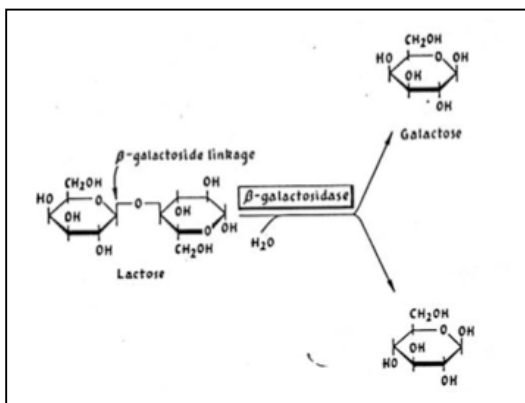
Phenotype: Lac⁻; what's observed

Genotype: *lacZ*⁻ or *lacY*⁻
refers to specific gene defect

Particular mutations = "alleles"
e.g., *lacZ343*

Early knowledge of lactose metabolism:

1. lactose cleaved by cells (=beta-galactosidase)
2. lactose taken into cells (=lactose permease)
3. induction (lactose addition increases lactose cleavage and uptake rates)



Questions answered genetically:

1. How many genes are required for lactose utilization?
2. What is the mechanistic "logic" of induction? (i.e., Does induction involve a positively- or negatively-acting regulatory factor?)

1. How many genes are required for lactose utilization?

A. Isolate hundreds of Lac⁻ mutants
(Lac⁻₁, Lac⁻₂, ...)

B. Determine how many complementation groups are defined by the Lac⁻ mutations.

B. Determine how many complementation groups (=genes) are defined by the Lac⁻ mutations.

General form of complementation test: Does lac⁻₁/lac⁻₂ strain grow on lactose or not?

Lac⁻ mutant isolation:

F⁻ lac⁺ cells



Mutagenize



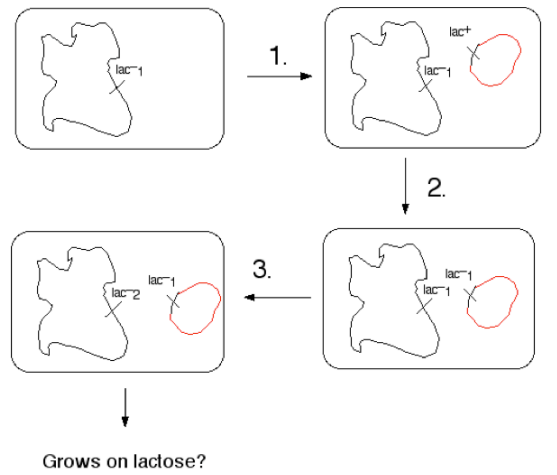
Plate cells for colonies and screen for mutants



lac⁻₁; lac⁻₂; etc

Complementation analysis of lac⁻ mutations

1. Transfer F'lac⁺ into Lac⁻ mutant (e.g., lac⁻₁)
2. Allow recombination to take place to transfer mutation (lac⁻₁) into F'lac
3. Conjugate F'lac carrying mutation (lac⁻₁) into second lac⁻ strain (e.g., lac⁻₂)
4. Examine whether F' lac⁻₁ / lac⁻₂ cells grow on lactose



	F'lac ⁻ ₁	F'lac ⁻ ₂	F'lac ⁻ ₃	F'lac ⁻ ₄
lac ⁻ ₁	-	-	+	+
lac ⁻ ₂	-	-	+	+
lac ⁻ ₃	+	+	-	-
lac ⁻ ₄	+	+	-	-
lac ⁻ ₅	-	-	+	+

Complementation group I = lac⁻_{1, 2, 5}

Complementation group II = lac⁻_{3, 4}

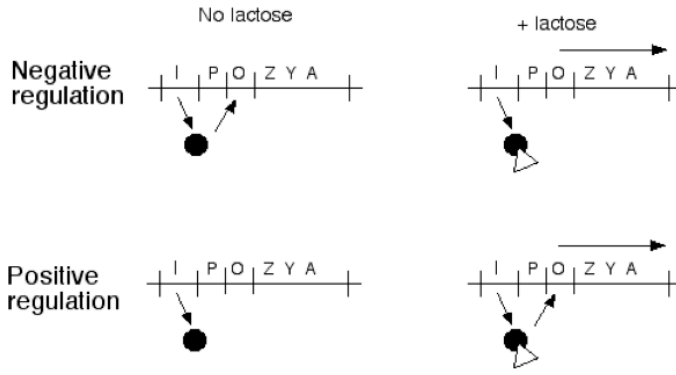
Coding assignments:

One set of mutants were beta-galactosidase⁻, transport⁺ (= lacZ⁻ lacY⁺)

Second set of mutants were beta-galactosidase⁺ transport⁻ (= lacZ⁺ lacY⁻)

2. Does induction involve a positively- or negatively-acting factor?

A. Alternative models: Negative vs positive



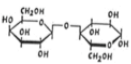
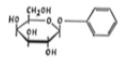
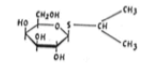
Models distinguished using constitutive mutants

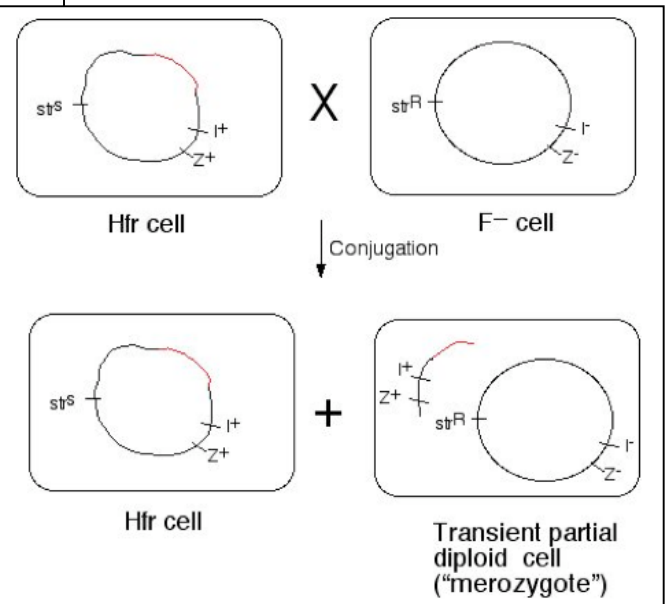
Negative regulation- predicts that the most common class of constitutive mutation would eliminate repressor

Positive regulation- predicts that the most common class of constitutive mutation would permanently activate activator

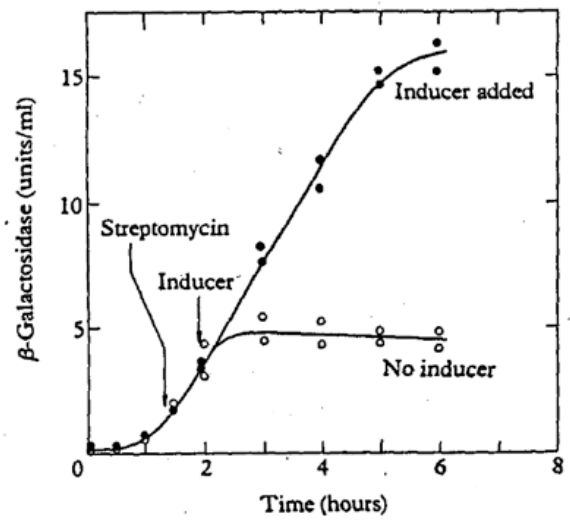
Steps:

1. Isolate constitutive mutants
2. Determine whether such mutations are dominant or recessive to wild-type

Substance	Cleaved by β -galactosidase	Induces <i>lac</i> operon transcription
Lactose 	+	+
Phenyl- β ,D-galactoside (PG) 	+	-
Isopropyl- β ,D-thiogalactoside (IPTG) 	-	+



Predictions for positive vs negative regulation (no inducer added)



Why are most common constitutive mutants $lacI^-$ rather than $lacO^-$?

How could one selectively isolate $lacO^-$ mutants?